49. Synthesis of Tenuilobine, a Bis-polyamine Alkaloid from *Oncinotis tenuiloba*, and Its Transamidation to Isotenuilobine

by Martin K.-H. Doll¹), Armin Guggisberg, and Manfred Hesse*

Organisch-chemisches Institut der Universität Zürich, Winterthurerstrasse 190, CH-8057 Zürich

(11.I.96)

From the leaves of Oncinotis tenuiloba STAPF, a novel polyamine alkaloid, tenuilobine (9), was isolated. This paper presents the synthesis of 9, as well as the base-catalyzed Zip reaction of 9, leading to the transamidation product isotenuilobine (10). The structure of 10 was further confirmed by 2D-NMR correlation spectroscopy. For analytical purposes, the bis-polyamines 9 and 10 were converted into their pentaacetyl derivatives 12 and 11, respectively, which were readily separable by reverse-phase HPLC.

1. Introduction. – Plants of the genus Oncinotis (Apocynaceae) are well-known for their content of polyamine alkaloids. Several different types of alkaloids have been isolated, ranging from simple monoacylated polyamines [1] to bi- or macrocyclic lactam alkaloids, all of them containing spermidine (1) as the basic component (Scheme 1). However, tenuilobine (9) is the first spermine derivative which was isolated from Oncinotis species [2]. Moreover, since 9 contains not only spermine (3) but spermidine (1) as well, this compound can be classified as a bis-polyamine alkaloid, a hitherto unknown subdivision of naturally occurring polyamine derivatives. In general, neither from plants [3] nor from animals [4], a substance was reported which contains two polyamine portions separated by an aliphatic moiety. The structure of tenuilobine (9) was elucidated by spectroscopic means and found to consist of hexadecanedioic acid being cross-linked with the secondary amino groups of spermidine (1) and spermine (3). To confirm these findings, we synthesized the alkaloid 9 as well as its unbranched isomer isotenuilobine (10), which could not be detected in the plant extract.

$$H_{2}N-(CH_{2})_{3}-N-(CH_{2})_{4}-NH_{2} \xrightarrow{a)} ZHN-(CH_{2})_{3}-N-(CH_{2})_{4}-NHZ \xrightarrow{H} H_{2}N-(CH_{2})_{3}-N-(CH_{2})_{4}-N-(CH_{2})_{3}-NHZ \xrightarrow{H} H_{2}N-(CH_{2})_{3}-N-(CH_{2})_{4}-N-(CH_{2})_{3}-NHZ \xrightarrow{H} H_{3} \xrightarrow{B} ZHN-(CH_{2})_{3}-N-(CH_{2})_{4}-N-(CH_{2})_{3}-NHZ \xrightarrow{H} H_{3} \xrightarrow{A} 4$$

0.1

Z = benzoyloxycarbonyl a) ZCN in CHCl₃, 23°. b) 2 equiv. ZCN in CHCl₃; 1 equiv. ZCN/Et₃N in CHCl₃, 23°.

¹) Part of the planned Ph. D. Thesis of M. K.-H. D., Universität Zürich.

2. Synthesis and Discussion. – Since hexadecanedioic acid had to be linked with two different polyamines, its monomethyl ester 5 was found to be a suitable starting material for this purpose. Unfortunately, all attempts to prepare 5 from the commercially available dimethyl ester by selective monohydrolysis were unsuccessful or resulted in very low yield. This problem could be circumvented by applying a two-step sequence using 16-hydroxyhexadecanoic acid as the starting material. *Fischer* esterification, followed by oxidation of the resulting methyl 16-hydroxyhexadecanoate with chromic acid, yielded the desired monomethyl ester 5 in 92% overall yield.

The polyamines spermidine (1) and spermine (3) were converted into their di- and triprotected benzyloxycarbonyl (Z) derivatives 2 and 4, respectively, prior to their use in the subsequent coupling reactions (*Scheme 1*). In general, the sterically less hindered NH_2 groups of spermidine (1) and related polyamines can be acylated selectively, when the reaction is carried out under kinetic control. Furthermore, the scope of the reaction seems to be limited to the use of sterically hindered carboxylic-acid derivatives; *e.g.* 3,4-dimethoxybenzonitrile [5] and 2-[(*tert*-butoxy)carbonyloxyimino]-2-phenylaceto-nitrile (Boc-ON) [6] have been used successfully for the preparation of N^1 , N^8 -diprotected spermidines²).

In our case, benzyl cyanoformate (ZCN) was chosen to prepare the corresponding N^1, N^8 -bis(Z)-protected spermidine **2**, since the final removal of the Z-protecting groups proceeds very cleanly in high yield by hydrogenolysis [7]. The original procedure of *Murahashi et al.* [5] [8] using ZCN in CH₂Cl₂ was unsuccessful due to the low solubility of spermidine (1). When CHCl₃ was used instead, the desired spermidine derivative **2** was obtained in 71 % yield after recrystallization. Moreover, we could extend the reaction to produce N^1, N^4, N^{12} -tris(Z)-protected spermine derivative **4** in high yield, when 3 equiv. of the reagent were added in two portions and in the presence of 3 equiv. of Et₃N.

Other reagents to produce N^1 , N^8 -bis(Z)-spermidine (2) have also been reported; *e.g.* 3-(benzyloxycarbonyl)-1,3-thiazolidine-2-thione generates 2 in 69% yield, when reacted with spermidine (1) [9]. Additionally, carbobenzoxyimidazole has been used for this purpose, affording 76% of 2 [10].

The most common procedure for amide formation from carboxylic acids and partially protected polyamines involves the *in situ* formation of the corresponding acid halide, which rapidly acylates the amine. This method has already been applied to the synthesis of related symmetrically cross-linked polyamines together with another procedure using phenyl bis(2-thioxo-1,3-thiazolidin-3-yl)phosphine oxide as the acyl-transfer agent [11]. Furthermore, the activated 4-nitrophenyl ester of the corresponding carboxylic acid can be used in more sensitive coupling reactions [12]. The method of *Mukaiyama* and coworkers who employed 1-methyl-2-chloropyridinium iodide as the coupling reagent [13] seemed most appropriate for our purposes. Thus, acylation of **2** with the monoester **5** furnished the intermediate **6** in 96% yield. After hydrolysis of the methyl-ester moiety in **6**, the resulting **7** was further cross-coupled with the protected spermine derivative **4** using *Mukaiyama*'s reagent at 23° to provide the all-Z-protected tenuilobine **8** in 91% yield. Final deprotection of **8** (H_2 /Pd) afforded tenuilobine (**9**) in 87% yield (*Scheme 2*).

²) For convenience, the following atom numbering is used in the text (for systematic names, cf. the Exper. Part): H₂NCH₂(1)CH₂CH₂(3)NH(4)CH₂CH₂CH₂CH₂(8)NH₂ and H₂NCH₂(1)CH₂CH₂(3)NH(4)CH₂CH₂CH₂CH₂(8)NH(9)CH₂CH₂CH₂(12)NH₂.



a) 1-Methyl-2-chloropyridinium iodide/Et₃N in CH₂Cl₂, reflux. b) MeOH/2N aq. NaOH, 23°. c) 1-Methyl-2-chloropyridinium iodide/Et₃N in CH₂Cl₂, 23°. d) 5% Pd/C, AcOH. e) KH/Propane-1,3-diamine (KAPA), 23°. f) Ac₂O/NaOAc, 23°.

The synthetic product 9 and the natural alkaloid were found to be identical in all respects, including their 2D-NMR spectra (¹H, ¹H TOCSY) [2].

3. Transamidation. – The isomerization of N-(3-aminopropyl)-amides of the general structure **13** leading to their counterparts **14** is known as the Zip reaction [14] [15] (Scheme 3). In general, strongly basic or acidic conditions are required, but the reaction proceeds also thermally [16]. If \mathbb{R}^1 and \mathbb{R}^2 of **13** constitute a ring, then ring enlargement



occurs. In this case, the analogous transamidation may also prove successful in the case of the corresponding N-(2-aminoethyl)- or N-(4-aminobutyl)-lactams, depending on the relative stability of the corresponding ring systems. However, it has been established that the reaction of the N-(4-aminobutyl) analogs proceeds much slower, when compared with compounds of type 13, due to the formation of the thermodynamically favored six-membered intermediate in the latter case as compared to the seven-membered counterpart.

Therefore, tenuilobine (9), when treated with base, would be expected to lead to isotenuilobine (10) with both polyamine moieties being linked at the primary amino groups of the propanediyl portion rather than at those of the butanediyl part (*Scheme 2*). To examine this expectation, 9 was treated with KAPA (KH/propane-1,3-diamine) at 23° (30 min), and, indeed, the structure of the resulting product was unambiguously established as 10 by 2D-NMR techniques.

With respect to C(10) (δ 180.44) in the ¹H,¹³C-HMBC spectrum of 10, cross-peaks were observed only for H–C(8) (δ 3.37), H–C(11) (δ 2.34), and H–C(12) (δ 1.66). The assignment of the signals was further supported by their chemical shifts. Additional ¹H,¹H-TOCSY experiments revealed cross-peaks between H–C(8), H–C(7) (δ 2.02), and H–C(6) (δ 3.16) as well as between H–C(11), H–C(12), and H–C(13–22) (δ 1.37), respectively, indicating the presence of the RCH₂CH₂CONH(CH₂)₃NHR' portion in the molecule as expected from the *Zip* reaction.

It should be noted that transamidation occurs even during storage of pure 9 at 4° (5 months), as it was also the case for pyridine solutions (4°, 4 weeks). Examination of such samples using TLC³) revealed the formation of an intermediate (*ca.* 20%, R_f 0.20) accompanied only by a trace of 10 (< 5%, R_f 0.16). Unchanged tenuilobine (9) was still predominant (*ca.* 80%, R_f 0.27). Although it is not clear whether the intermediate consists of a mixture of the two possible partly transamidated isomers of 9, or if it is exclusively one of them, it can be stated, that the transamidation $9 \rightarrow 10$ proceeds stepwise rather than synchronously.

Surprisingly, the ESI-MS/MS spectra of 9 and 10 are identical with respect to the observed fragment ions, except their relative intensities. This might be due to a thermally

³) Polygram^{*} SIL N-HR/UV₂₅₄, Macherey-Nagel; CHCl₃/MeOH/25% aqueous NH₃ solution 6:3:1.

promoted rearrangement of 9 during MS measurement, in the same manner as discussed above. Unfortunately, no direct comparison of the NMR spectra of 9 and 10 was possible, since the latter was hardly soluble in (D_5) pyridine, and D_2O had to be used as solvent⁴).

For HPLC purposes, 9 and its transamidated isomer 10 were converted into their pentaacetyl derivatives 12 and 11, respectively, which could be easily distinguished using reverse-phase conditions (*cf. Exper. Part*). Additionally, coinjection of 12 together with the pentaacetyl derivative of the alkaloid isolated from natural sources demonstrated the identity of the samples, and the absence of 11 in the acetylated extract showed that 10 is not a natural product. This has been already shown using a different system of chromatography [2].

The financial support from the *Swiss National Science Foundation* is greatly appreciated. Thanks are due to Dr. D. Nanz and Mr. D. Rentsch for recording 2D-NMR spectra and helpful discussions, and Dr. E. Reder and Mr. L. Bigler for measuring the mass spectra. Technical assistance by Ms. C. Sprenger is gratefully acknowledged.

Experimental Part

General. For the slow addition of ZCN, an infusion pump (*Precidor 5003, Infors AG*, Switzerland) was used. Column chromatography (CC): *Merck* silica gel 60 (40–60 μ). IR (cm⁻¹): in CHCl₃, *Perkin-Elmer 781*. ID-NMR Spectra: *Bruker AMX-600, Bruker AM 400*, and *Bruker ARX 300* with chemical shifts δ in ppm, coupling constants *J* in Hz, using the appropriate solvent as internal standard. Unless stated otherwise, ¹H-NMR: at 300.1 MHz. ¹³C-NMR: at 75.5 MHz in CDCl₃. 2D-NMR (D₂O): *Bruker AMX-600*; TOCSY (mixing time 150 ms) and HMBC (*J* selected for 9 Hz) [19]. ESI-MS: *Finnigan TSQ 700* mass spectrometer; for MS/MS, Ar (2.5 mtorr) was used as collision gas.

Methyl 16-Hydroxyhexadecanoate. An ethereal soln. of CH₂N₂ (freshly prepared) was added dropwise to a stirred soln. of 16-hydroxyhexadecanoic acid (1.0 g, 36.7 mmol) in MeOH (25 ml), until a slightly yellow color persisted. Evaporation of the solvent *in vacuo* left 1.0 g (95%) of the product. M.p. 54.7–55.1° (recrystallized from MeOH/cyclohexane/pentane). ¹H-NMR: 3.59 (*s*, Me); 3.57 (*t*, J = 6.7, CH₂OH); 2.23 (*t*, J = 7.5, CH₂CO₂Me); 1.57–1.30 (*m*, 2 CH₂); 1.19 (*s*-like *m*, 11 CH₂).

15-(Methoxycarbonyl)pentadecanoic Acid (5). An aq. soln. of chromic acid (9.41 ml, prepared from 2.085 g of CrO₃, 30 ml of H₂O, and 8.7 ml of H₂SO₄) was added over 20 min to a stirred soln. of methyl 16-hydroxyhexadecanoate (907.3 mg, 3.2 mmol) in acetone (50 ml) and stirring was continued for 3 h. After addition of H₂O (15 ml), the acetone was evaporated *in vacuo*, Na₂S₂O₅ was added and the aq. soln. extracted with Et₂O. The combined Et₂O layers were re-extracted once (H₂O), dried (Na₂SO₄), and evaporated: 926.8 mg (97%) of 5. M.p. 68.7–71.9°. ¹H-NMR: 3.66 (*s*, Me); 2.34 (*t*, *J* = 7.5, CH₂CO₂H); 2.30 (*t*, *J* = 7.6, CH₂CO₂Me); 1.67–1.58 (*m*, 2 CH₂); 1.35–1.25 (*m*, 10 CH₂).

Benzyl N- {[8-(Benzyloxycarbonyl)amino]-4-azaoctyl}carbamate (2). A soln. of spermidine (1) (3.56 g, 24.5 mmol) in CHCl₃ (230 ml; purified on *ICN* Alumina **B**, activity 1) was stirred under N₂ at 23°. During 6 h, a soln. of PhCH₂OCOCN (7.9 g, 49.02 mmol) in CHCl₃ (80 ml) was added, and stirring was continued for 16 h. Evaporation of the solvent and recrystallization of the residue (acetone/hexane) yielded 7.17 g (71%) of **2**. M.p. 105.9–107.5°. IR: 3450m, 2940m, 1712s, 1511s, 1455w, 1260m, 1025w. ¹H-NMR: 7.27–7.19 (*m*, 10 arom. H); 5.49, 5.20 (br. 2s, 2 NHCO); 5.00 (s, 2 PhCH₂); 3.21–3.15, 3.11–3.07 (2 q-like m, 2 CH₂NHCO); 2.57 (t, *J* = 6.4, CH₂NH); 2.52–2.48 (t, *J* = 6.2, CH₂NH); 1.61–1.52 (quint.-like m, CH₂); 1.48–1.39 (m, 2 CH₂). ¹³C-NMR: 155.47 (s, 2 CO); 135.72 (s, 2 C_{ipso}); 127.47, 127.02 (2d, 10 arom. CH); 65.50 (t, 2 PhCH₂); 48.32, 46.74, 39.90, 38.89, 28.54, 26.77, 26.30 (7t, 7 CH₂). ESI-MS/MS (of *m*/z 414.5, –20 eV): 414.4 (15), 306.5 (3), 181.2 (7), 148.3 (13), 91.4 (100, [C₆H₄CH₂]⁺), 72.6 (4).

Dibenzyl N, N-[4-(Benzyloxycarbonyl)-4,9-diazadodecane-1,12-diyl]bis[carbamate] (4). A soln. of spermine (3; 5.0 g, 24.7 mmol) in CHCl₃ (250 ml, purified as described for 2) was stirred under Ar at 23°. To this soln.

⁴) Although the NMR spectra of 10 were well resolved in D₂O, attempts to record the spectra of 9 in the same solvent resulted in extremely broad signals.

PhCH₂OCOCN (8.0 g, 49.6 mmol) in CHCl₃ (120 ml) was added during 3 h and stirring was continued overnight. After the addition of Et₃N (7.48 g, 74.05 mmol), additional PhCH₂OCOCN (4.18 g, 25.94 mmol) in CHCl₃ (70 ml) was added within 2 h and stirring was continued for 20 h. Evaporation of the solvent and CC of the residue (CHCl₃/MeOH/25% aq. NH₃ soln. 90:10:0.5) gave 12.74 g (85%) of 4. Brownish, viscous oil. IR: 3450m, 2940m, 1710s, 1512s, 1240s, 1140m, 695m. ¹H-NMR: 7.34–7.28 (*m*, 15 arom. H); 5.63–5.56 (br. *m*, 2 NHCO); 5.10–5.08 (*d*-like *m*, 3 PhCH₂); 3.27–3.16 (br. *m*, 4 CH₂NHCO); 2.62–2.52 (br. *m*, 2 CH₂NH); 1.75–1.42 (br. *m*, 4 CH₂). ¹³C-NMR (100 mg in 0.6 ml): 156.60, 156.45 (2s, 3 CO); 136.69 (s, 3 C_{ipso}); 128.45, 128.38, 128.19, 127.92, 127.77, 126.80 (6d, 15 arom. CH); 67.07, 66.40 (2t, 3 PhCH₂); 49.09, 47.32, 46.58, 44.14, 39.48, 38.15, 37.69, 29.18, 28.03, 26.70, 26.19, 25.59 (12t, CH₂). ESI-MS (MeOH + 1% TFA): 1211.9 (12, $[2M + 1]^+$), 606.6 (100, $[M + 1]^+$). ESI-MS/MS (of *m*/z 606.3, –20 eV): 606.5 (79), 562.3 (9), 353.7 (9), 263.7 (43), 112.2 (11), 91.1 (100, $[C_6H_5CH_2]^+$), 84 (9).

 $Methyl = 15 - \{\{\{4-[(Benzyloxycarbonyl)amino]butyl\}\{3-[(benzyloxycarbonyl)amino]propyl\}amino\}carbon$ yl pentadecanoate (6). To a suspension of 1-methyl-2-chloropyridiniumiodide (307 mg, 1.2 mmol) in abs. CH₂Cl₂ (10 ml) were added 5 (300 mg, 1.0 mmol) and Et₃N (243 mg, 2.4 mmol), and the mixture was stirred under Ar at 23°. After dropwise addition of a soln. of 2 (517 mg, 1.3 mmol) in CH₂Cl₂ (5 ml), the mixture was refluxed for 1.5 h and stirred at 23° overnight. The solvent was evaporated, the residue taken up in CH_2Cl_2 (50 ml), and the soln. extracted with 0.5N aq. HCl soln. and with brine. Drying of the org. layer (Na₂SO₄), evaporation of the solvent, and CC (CH₂Cl₂/MeOH 49:1) of the residue yielded 670.4 mg (96%) of 6. IR: 3450m, 2925s, 2855m, 1718s, 1625s, 1510s, 1240s, 1140w, 1015w. ¹H-NMR (400.1 MHz): 7.27-7.21 (m, 10 arom. H); 5.77 (br., NH); 5.08-5.00 (d-like m, 2 PhCH₂); 4.93 (br., NH); 3.58 (s, Me); 3.31-3.28 (m, CH₂NHCO); 3.21-3.11 (m, (CH₂)₂NCO); 3.05-3.00 (m, CH₂NHCO); 2.24–2.17 (*m*, CH₂CO₂Me, CH₂CON); 1.67–1.42 (br. *m*, 5 CH₂); 1.20–1.17 (*m*, 10 CH₂). ¹³C-NMR (100.6 MHz): 174.19, 173.55 (2s, CO₂Me, CON); 156.46 (s, 2 NHCO); 136.77 (s, 2 C_{ipso}); 128.39, 128.28, 128.00, 127.92, 127.84 (5d, 10 arom. CH); 66.52, 66.21 (2t, 2 PhCH₂); 51.23 (s, Me); 47.24, 45.33, 45.15, 42.12, 40.29, 38.52, 37.61, 33.95, 32.95, 29.46, 29.36, 29.08, 28.98, 27.72, 27.34, 27.08, 25.95, 25.43, 25.31, 24.80, 24.78 (21t, 21 CH₂). ESI-MS (CHCl₃/MeOH 2:1 + 5% TFA): 1392.5 (36, $[2M + 1]^+$), 718.6 (23, $[M + Na]^+$), 696.5 (100, $[M + 1]^+$), 588.6 (26), 480.5 (16). ESI-MS/MS (of m/z 696.5, -15 eV): 696.5 (20), 652.4 (42), 588.5 (27), 544.3 (37), 480.5 (100), 448.3 (46), 437.0 (29), 198.2 (45).

15 - {{{4 - [(Benzyloxycarbonyl)amino]butyl}3 - [(benzyloxycarbonyl)amino]propyl}amino]carbonyl}pentadecanoic Acid (7). To a soln. of 6 (620.4 mg, 0.89 mmol) in MeOH (15 ml) was added 2N aq. NaOH soln. (2.4 ml) at 23°. After stirring for 24 h, acidification (pH 5) with 2N aq. HCl soln. and evaporation of the solvents, the residue was taken up in CHCl₃ and filtered to remove NaCl. Evaporation of the filtrate yielded 7 in quant. yield. Colorless lac. IR: 3450m, 2930s, 2860m, 1711s, 1625s, 1510s, 1235s, 1140w, 1015w. ESI-MS (MeOH): 720.8 (12, $[M + K]^+$), 704.8 (100, $[M + Na]^+$).

Dibenzyl N,N- { $26 - (3 - Aminopropyl) - 4 - (benzyloxycarbonyl) - 9 - {3 - [(benzyloxycarbonyl)amino]propyl} - 10,25-dioxo-4,9,26-triazatricosane-1,30-diyl}bis[carbamate] (8). A mixture of 7 (627.3 mg, 0.92 mmol), 1-methyl-2-chloropyridinium iodide (282.0 mg, 1.1 mmol), and Et₃N (223.0 mg, 2.2 mmol) in CH₂Cl₂ (30 ml) was stirred under Ar at 23°. A soln. of 4 (690 mg, 1.14 mmol) in CH₂Cl₂ (10 ml) was added dropwise and stirring was continued for 16 h. The solvent was evaporated, CH₂Cl₂ (100 ml) was added and the soln. washed with 0.5N aq. HCl soln. (2 × 50 ml) and 5% aq. Na₂CO₃ soln. Drying (Na₂SO₄) and evaporation of the org. layer left 1.4 g of crude material, which afforded, after CC (CHCl₃/MeOH 98:2), 1.065 g (91%) of colorless, oily 9. IR: 3450w, 2930m, 2860w, 1715s, 1628m, 1512s, 1260s, 1090m, 1015m. ¹H-NMR: 7.33-7.28 (m, 25 arom. H); 5.10-5.08 (m, 5 PhCH₂); 3.38-3.09 (br. m, 10 CH₂N); 2.28-2.23 (m, 2 CH₂CON); 1.71-1.48 (br. m, 9 CH₂); 1.26-1.24 (m, 10 CH₂); 128.50, 128.47, 128.41, 127.95 (4d, 25 arom. CH); 6.7.31, 66.67, 66.37 (3t, 5 PhCH₂); 47.38, 46.47, 45.37, 45.16, 44.26, 42.27, 40.40, 38.60, 37.74, 33.07, 29.61, 29.50, 28.91, 27.84, 27.48, 26.06, 25.58 (17t, CH₂). ESI-MS (CHCl₃/MeOH 2:1): 1292.2 (100, [M + Na]⁺), 657.4 (13, <math>\frac{1}{2}[M + 2Na]^{2+}$).

 N^{1} -(4-Aminobutyl)- N^{1} , N^{16} -bis(3-aminopropyl)- N^{16} -{4-[(3-aminopropyl)amino]butyl}hexadecanediamide (= Tenuilobine; 9). To a soln. of 8 (463 mg, 0.36 mmol) in AcOH (100 ml) was added 5% Pd/C (150 mg), and the mixture was hydrogenated in a *Parr* apparatus (3.5 bar, 16 h). Evaporation and CC of the residue (CHCl₃/MeOH/ 25% aq. NH₃ soln. 5:3:1) gave 190 mg (87%) of 9. Colorless oil. ¹³C-NMR (150.9 MHz, (D₅)pyridine): 172.53, 172.34 (2s, 2 CO); 50.07, 49.94, 48.41, 48.35, 47.79, 45.74, 45.70, 42.97, 42.93, 42.29, 42.22, 41.00, 39.74, 39.65, 34.14, 34.00, 33.29, 33.24, 32.16, 31.48, 31.25, 29.98, 29.92, 29.88, 27.94, 27.74, 27.39, 26.98, 26.19, 26.07, 25.72, (32*t*, CH₂). For additional spectroscopic data *cf.* [2].

N¹-(4-Acetamidobutyl)-N¹, N¹⁶-bis(3-acetamidopropyl)-N¹⁶-{4-[(3-acetamidopropyl)(acetyl)amino]butyl}hexadecanediamide (12). Acetylation of 9 was performed as described in [2]. ¹³C-NMR (70 mg in 0.6 ml): 173.77, 173.66, 172.97 (3s, 2 CH₂CON); 171.03, 170.84, 170.66, 170.40, 170.32 (5s, 5 MeCO); 48.17, 47.40, 46.41, 44.73, 42.34, 42.25, 38.60, 36.88, 35.99, 35.90, 32.99, 29.40, 29.31, 29.12, 27.29, 26.90, 26.29, 26.07, 25.78, 25.48, 25.36, 24.88, 23.14, 22.93, 21.31 (25t, CH₂). For additional spectroscopic data see [2].

1,38-Diamino-5,9,26,30,35-pentaazaoctatriacontane-10,25-dione (= *Isotenuilobine*; **10**). A suspension of KH (600 mg, 15 mmol) in propane-1,3-diamine (5 ml) was stirred at 23° under Ar, until H₂ evolution ceased. A soln. of **9** (51.4 mg, 0.086 mmol) in a small amount of propane-1,3-diamine was added dropwise and stirring was continued for 30 min. The mixture was carefully dropped into 0.3N aq. HCl soln. (15 ml) and the solvents were removed *in vacuo*. The residue was taken up in H₂O, basified (K₂CO₃), and extracted with CHCl₃. Drying of the org. layer (Na₂SO₄), evaporation, and CC (CHCl₃/MeOH/25% aq. NH₃ soln. 5:3:1) yielded 42.6 mg (83%) of **10**. Colorless oil. IR (KBr): 3335s, 2920s, 2485m, 1652s, 1531s, 1460s, 1405s, 1265w, 1170w, 1055w, 725w. ¹H-NMR (600.1 MHz, D₂O): 3.37 (*t*, *J* = 6.7, CH₂NHCO); 3.28–3.25 (*t*-like *m*, CH₂); 1.91–1.83 (*m*, 4 CH₂); 1.68–1.65 (*m*, 2 CH₂); 1.38–1.36 (*m*, 10 CH₂). ¹³C-NMR (150.9 MHz, D₂O): 180.44 (*s*, 2 CO); 50.01, 49.96, 49.91, 48.15, 48.13, 47.54, 41.80, 39.56 (8*t*, 8 CH₂); 25.78 (*t*, 2 CH₂). ESI-MS (MeOH/H₂O 1:1): 598.9 (31. [*M* + 1]⁺), 300.1 (61, ½[*M* + 2]²⁺), 200.3 (100, ½[*M* + 3]³⁺). ESI-MS/MS (of *m*/z 68.9, -25 eV): 599.1 (66), 581.1 (12), 525.4 (35), 508.1 (37), 454.1 (29), 436.7 (38), 383.1 (35), 365.2 (45), 307.6 (15), 129.2 (100), 112.1 (26), 72.4 (7).

5,30,35-Triacetyl-1,38-bis(acetylamino)-5,9,26,30,35-pentaazaoctatriacontane-10,25-dione (11). A sample of 10 was peracetylated as described in [2]. ESI-MS (MeOH): 830.9 (52, $[M + Na]^+$), 427.0 (100, $\frac{1}{2}[M + 2Na]^{2+}$).

HPLC-Separation of 11 and 12. *MN Nucleosil* 100-5C₈ (200-8-4), MeOH/H₂O (58:42; flow 1.5 ml/min), UV detection at 225 nm: t_R 22.8 min for 11 and 25.4 min for 12 with $R \approx 0.87$.

REFERENCES

- [1] M. K.-H. Doll, A. Guggisberg, M. Hesse, Helv. Chim. Acta 1994, 77, 1229.
- [2] M. K.-H. Doll, A. Guggisberg, M. Hesse, Heterocycles 1996, 42, 319.
- [3] A. Guggisberg, M. Hesse, in 'The Alkaloids', Ed. A. Brossi, Academic Press Inc., New York, 1983, Vol. 22, p. 85.
- [4] A. Schäfer, H. Benz, W. Fiedler, A. Guggisberg, S. Bienz, M. Hesse, in 'The Alkaloids', Ed. G.A. Cordell, A. Brossi, Academic Press Inc., San Diego, 1994, Vol. 45, p. 1.
- [5] S.-I. Murahashi, T. Naota, N. Nakajima, Chem. Lett. 1987, 879.
- [6] W. J. Fiedler, Ph. D. Thesis, University of Zürich, 1992.
- [7] M. Bergmann, L. Zervas, Chem. Ber. 1932, 65, 1192.
- [8] S.-I. Murahashi, T. Naota, Synthesis 1993, 433.
- [9] Y. Nagao, T. Miyasaka, Y. Hagiwara, E. Fuyita, J. Chem. Soc., Perkin Trans. 1 1984, 183.
- [10] S.K. Sharma, M.J. Miller, S.M. Payne, J. Med. Chem. 1989, 32, 357.
- [11] G. Sosnovsky, J. Lukszo, Z. Naturforsch., B, 1994, 49, 1580.
- [12] J. R. Goodnow, K. Konno, M. Niwa, T. Kallimopoulos, R. Bukownik, D. Lenares, K. Nakanishi, Tetrahedron 1990, 46, 3267.
- [13] E. Bald, K. Saigo, T. Mukaiyama, Chem. Lett. 1975, 1163.
- [14] U. Kramer, A. Guggisberg, M. Hesse, H. Schmid, Angew. Chem. Int. Ed. 1978, 17, 200.
- [15] U. Kramer, A. Guggisberg, M. Hesse, H. Schmid, Helv. Chim. Acta 1978, 61, 1342.
- [16] A. Guggisberg, B. Dabrowski, U. Kramer, C. Heidelberger, M. Hesse, H. Schmid, Helv. Chim. Acta 1978, 61, 1039.
- [17] U. Kramer, A. Guggisberg, M. Hesse, H. Schmid, Angew. Chem. Int. Ed. 1977, 16, 861.
- [18] E. Stephanou, A. Guggisberg, M. Hesse, Helv. Chim. Acta 1979, 62, 1932.
- [19] A. Bax, M. F. Summers, J. Am. Chem. Soc. 1986, 108, 2093.